

1 **Article Type: Perspective**

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3 **Recognizing taste: coding patterns along the neural axis in mammals**

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21

22 **Abstract**

23 The gustatory system encodes information about chemical identity, nutritional value, and concentration
24 of sensory stimuli before transmitting the signal from taste buds to central neurons that process and
25 transform the signal. Deciphering the coding logic for taste quality requires examining responses at each
26 level along the neural axis - from peripheral sensory organs to gustatory cortex. From the earliest single
27 fiber recordings, it was clear that some afferent neurons respond uniquely, others to stimuli of multiple
28 qualities. There is frequently a “best stimulus” for a given neuron, leading to the suggestion that taste
29 exhibits “labeled line coding”. In the extreme, a strict “labeled line” requires neurons and pathways
30 dedicated to single qualities (e.g. sweet, bitter, etc.). At the other end of the spectrum, “across-fiber”,
31 “combinatorial”, or “ensemble” coding requires minimal specific information to be imparted by a single
32 neuron. Instead, taste quality information is encoded by simultaneous activity in ensembles of afferent
33 fibers. Further, “temporal coding” models have proposed that certain features of taste quality may be
34 embedded in the cadence of impulse activity. Taste receptor proteins are often expressed in non-
35 overlapping sets of cells in taste buds apparently supporting “labeled lines”. Yet, taste buds include both
36 narrowly- and broadly-tuned cells. As gustatory signals proceed to the hindbrain and on to higher
37 centers, coding become more distributed, and temporal patterns of activity become important. Here, we
38 present the conundrum of taste coding in the light of current electrophysiological and imaging techniques
39 at several levels of the gustatory processing pathway.

40

41 **Keywords:** gustatory coding, taste quality, taste bud, geniculate ganglion, nucleus of solitary tract,
42 gustatory cortex

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44

45 **Introduction**

46 All sensory systems must address the problem of conveying information about the quality, intensity, and
47 location of sensory stimulation from peripheral receptors to the brain. For both olfaction and taste, stimuli
48 can be chemically diverse. The olfactory system is known to encode this chemical diversity, in part,
49 through the use of hundreds of molecular receptors with overlapping receptive ranges. Olfactory signals
50 from peripheral neurons are carried on circuits that exhibit convergence and distributed patterns at
51 different stages along the neural axis to encode odor recognition and discrimination (Laurent 2002; Nara
52 *et al.* 2011; Nunez-Parra *et al.* 2014; Srinivasan and Stevens 2018). The gustatory system, which serves
53 to detect nutrients, minerals, and toxins, also identifies diverse chemical structures across broad
54 concentration ranges. The logic of how the mammalian gustatory system encodes information on
55 chemical identity, i.e. quality coding, is the subject of active investigation using a variety of experimental
56 approaches and resulting in competing models of taste coding. The present review examines some of
57 the evidence, interpretations and controversies regarding gustatory quality coding.

58 Most research on taste quality coding focuses on discriminating “sweet” (for example sugars), “salty”
59 (Na^+ salts), “sour” (acids) and so forth. Labeled line coding posits that quality-specific taste receptor
60 cells (TRCs) (for example “sweet”-specific) synapse only with primary sensory afferent(s) that are
61 dedicated to that same quality. This, then, establishes a dedicated transmission line from the taste bud
62 cell to the brain that is “labeled” for a single quality. According to this coding, the different transmission
63 lines (“sweet”, “salty”, “sour”, etc.) are separate, distinct, and parallel. The sensory afferent neurons are
64 all highly tuned to transmit one given quality. They are all “specialists” for a given quality.

65 In contrast, combinatorial coding allows more flexibility in the responses of primary afferent fibers. Thus,
66 a given taste compound can elicit impulses in an ensemble of several primary afferent fibers, each of
67 which varies in their response profiles. That is, some fibers might be “sweet-best”, others might be “salt-
68 best”; they respond robustly to sugars or Na^+ salts, respectively, while retaining weaker responses to
69 other tastes (“specialists”). Other fibers in the ensemble may respond quite broadly to many different
70 taste compounds with no strong preference (“generalists”). However, when activated by a specific taste
71 compound, the entire ensemble of afferent fibers generates a particular combinatorial signal that
72 identifies that stimulus. Collectively, the combination of specialists and generalists, not any individual
73 sensory afferent axon on its own, transmits the information about taste quality. Temporal coding conveys
74 information in the pattern of impulses in individual primary sensory afferents. Different taste stimuli may
75 elicit different patterns of action potentials in afferent fibers, which might lead to differential
76 excitation/inhibition of neurons in the CNS.

77 For theorists, both models present a dilemma: how do multi-sensitive cells convey an unambiguous
78 message that identifies taste quality? The labeled line and across-neuron pattern theories share the
79 notion that spikes are integrated over time, and ignore the dynamics of firing rate changes that occur
80 during a taste response. These dynamic aspects of the response may also carry taste information, a
81 form of signaling called “temporal coding”.

82 The origins of labeled line coding in the sensory nervous system might be said to come from René
83 Descartes, who, in his classical drawing of the innocent cherub toasting his toes (Descartes 1664, p.
84 27), clearly outlined a labeled line (here, for painful heat) from peripheral sensory organ to the brain
85 (Roper 2014). However, the first explicit statements of labeled line coding were by Sir Charles Bell (1811;
86 see Bell and Shaw 1868), and Johannes Müller (1835), who coined the concept, Law of Specific Nerve
87 Energies (LOSNE), according to which “each type of sensory nerve ending, however stimulated
88 (electrically, mechanically, etc.), gives rise to its own specific sensation; moreover, each type of
89 sensation depends not upon any special character of the different nerves but upon the part of the brain
90 in which their fibers terminate” (Müller 1836). Since then, it has become clear that each modality is
91 indeed “labeled” insofar as touch, temperature, taste, olfaction, vision, hearing, and so forth are each
92 transmitted along separate neural pathways. The question, now, is whether such “labeling” extends to
93 different qualities *within* a sensory modality, such as red versus green color, rose versus geranium scent,
94 or sweet versus salty taste. That is the crux of the current debate. In certain sensory systems, such as
95 vision and olfaction, the answer is clearly “no”; colors and odors unarguably display combinatorial quality
96 coding.

97 In this review, we examine the evidence, primarily derived from electrophysiological and imaging studies
98 at different levels of the taste system, of the responses of receptors and neurons to stimuli representing
99 different taste qualities. We discuss what the responses at each level suggest about the logic of coding
100 taste quality.

101

102 **The detectors: coding taste quality in taste bud cells**

103 A strict peripheral labeled line coding for taste qualities (sweet, salty, sour, etc.) has been strongly
104 promoted by some researchers (Barretto *et al.* 2015; Chen *et al.* 2011b; Yarmolinsky *et al.* 2009). The
105 strongest evidence for such a hard-wired logic for taste quality coding comes from the observation that
106 taste bud cells express primarily or only one type of taste receptor. Some cells express a few to several
107 members of the Tas2R family of receptors which are activated by bitter-tasting compounds (Behrens *et al.*
108 2007; Mueller *et al.* 2005). Other TRCs may express heterodimeric Tas1R family receptors, which
109 are activated by either sweet- or umami-tasting compounds (Dando *et al.* 2012; Nelson *et al.* 2002;

110 Nelson *et al.* 2001). Yet other cells are dedicated for sour taste sensing (Huang *et al.* 2006). However,
 111 some fraction of taste cells do express taste receptors for more than one quality (Dando *et al.* 2012).
 112 The relatively non-overlapping pattern of receptor expression led to the proposal that, similar to insects,
 113 mammals use a hard-wired logic for coding taste quality (Yarmolinsky *et al.* 2009). That is for example,
 114 Tas2R-expressing TRCs, when stimulated, activate a dedicated subset of afferent fibers which would
 115 encode the bitter taste quality. Other dedicated TRCs and nerve fibers would convey sweet and so on.
 116 This taste quality-dedicated TRCs constitute the beginning of a labeled line for “bitter” or “sweet”,
 117 respectively, that is maintained along the taste axis to the gustatory cortex.

118 The question is how well do the responses of individual taste bud cells mirror the seemingly
 119 compartmentalized, non-overlapping pattern of expression of the various taste receptors. The taste
 120 quality sensitivity and selectivity of specific populations of taste bud cells have been examined through
 121 both, electrophysiological and Ca^{2+} imaging methods (Tomchik *et al.* 2007; Yoshida *et al.* 2009; Yoshida
 122 *et al.* 2018), using several distinct *ex vivo* preparations. Using the combination of transgenically identified
 123 taste bud cell types and apical stimulation with a variety of taste stimuli, the response profiles of taste
 124 bud cell types have been studied electrophysiologically (Yoshida *et al.* 2009) and via Ca^{2+} imaging
 125 (Caicedo *et al.* 2002; Tomchik *et al.* 2007). Very consistently, Type II cells respond best to sweet, bitter
 126 or umami taste stimuli. “Bitter-best” taste cells are the most narrowly tuned and respond almost
 127 exclusively to bitter compounds (Yoshida *et al.* 2009b). In contrast, some “sweet-best” TRCs are more
 128 broadly tuned such that, in addition to sucrose, some also respond to salt (NaCl) and/or umami stimuli
 129 (monosodium glutamate, MSG). Type III cells from fungiform taste buds consistently respond to acid
 130 (sour) stimuli, and each cell typically responds to multiple acids (citric, acetic or HCl). Thus, tuning,
 131 measured in the electrical responsivity of cells from fungiform taste buds (Yoshida *et al.* 2009), is
 132 generally similar to that measured by the Ca^{2+} responses of Type II cells from mouse circumvallate taste
 133 buds (Tomchik *et al.* 2007). Further, in both studies, responses to acids were limited to Type III cells.

134 Type III cells in mouse fungiform papillae fell into 2 groups with approximately 75% responding only to
 135 acids, the rest being broadly tuned, with responses to salty, umami, and/or bitter stimuli in addition to
 136 acids. This observation differed conspicuously the Ca^{2+} imaging study which reported that all or most
 137 Type III cells in mouse circumvallate taste buds were both sour-responsive and broadly tuned (Tomchik
 138 *et al.* 2007). Whether these differences are attributable to differences in methodology or in the taste bud
 139 fields examined (fungiform vs. circumvallate) remains to be determined.

140 Another question that has been explored electrophysiologically in mouse fungiform taste bud cells is
 141 how diverse stimuli that produce similar taste perception are represented in the initial receptor cells. For
 142 example, many sugars (sucrose, fructose, etc.), artificial sweeteners (saccharin, sucralose, etc.) and
 143 certain proteins (Monellin, Thaumatin, Brazzein, etc.) all elicit sweet taste. Similarly, there are numerous

chemically diverse compounds, all of which elicit bitter taste. To test whether TRCs respond identically to diverse stimuli of a given quality (for example “bitter”) or can discriminate among perceptually similar compounds, responses were recorded to a battery of bitter-tasting compounds (Yoshida *et al.* 2018). Type II TRCs from fungiform and circumvallate taste buds showed considerable heterogeneity in their responses to this battery of bitter chemicals. Some bitter stimuli elicited responses in 5-8 times as many taste cells as did other bitter compounds. That is, taste compounds that are perceived as having similar taste may produce very different patterns of activation among taste bud cells

Yoshida *et al.* (2018) also demonstrated that bitter-sensitive cells as a population displayed considerable heterogeneity. When tested with 10 bitter compounds, some were selective for only a single stimulus while others responded broadly to as many as 9 of the 10 stimuli tested. Such heterogeneous responses among bitter-sensitive taste cells had also been demonstrated using functional imaging of rat and mouse circumvallate taste bud cells (Caicedo *et al.* 2002; Caicedo and Roper 2001). The family of bitter taste receptors Tas2Rs, includes ~35 diverse members and each of these Tas2Rs is activated by a different complement of bitter compounds (Lossow *et al.* 2016). In both, human and mouse, some Tas2rs are narrowly tuned and others that can be activated by large numbers of bitter tasting compounds (Lossow *et al.* 2016; Meyerhof *et al.* 2010). Thus, the selectivity of bitter sensitive TRCs would be defined by the expression of different combinations of Tas2Rs.

All molecular receptors for bitter tastants, Tas2Rs, were reported to be co-expressed in some TRCs with the interpretation that discrimination among bitter stimuli could not occur (Adler *et al.* 2000). More comprehensive analyses showed that only limited numbers of Tas2Rs are expressed per TRC, and in various combinations (Behrens *et al.* 2007; Matsunami *et al.* 2000). The electrophysiological and Ca²⁺ imaging results above also demonstrate that the initial hypothesis (Mueller *et al.* 2005) for how bitter taste quality is coded in the periphery was likely incorrect. Combinatorial expression of Tas2Rs in individual TRCs could, in principle, form a basis for discriminating among different bitter compounds, but it is unclear whether such discrimination exists along the taste neural axis or even behaviorally.

Taken together, electrophysiological and Ca²⁺ imaging data indicate that taste buds contain many taste receptor cells dedicated to detect one of 5 basic taste qualities. These may provide the basis for discrimination across basic taste qualities. However, taste buds also contain TRCs that respond to multiple taste qualities (Caicedo *et al.* 2002; Tomchik *et al.* 2007; Yoshida *et al.* 2009). These multiply-responsive cells may reflect information processing (divergence and convergence of signals) that occurs within taste buds via cell-cell synaptic interactions (Chaudhari 2014; Dando and Roper 2009; Huang *et al.* 2009; Huang *et al.* 2007). Moreover, some taste cells express multiple types of taste receptors. For instance, a subset of taste cells expresses all three T1R subunits and responds to both sweet and umami compounds (Dando *et al.* 2012; Kuschara *et al.* 2013). Whether broadly tuned TRCs serve a distinct

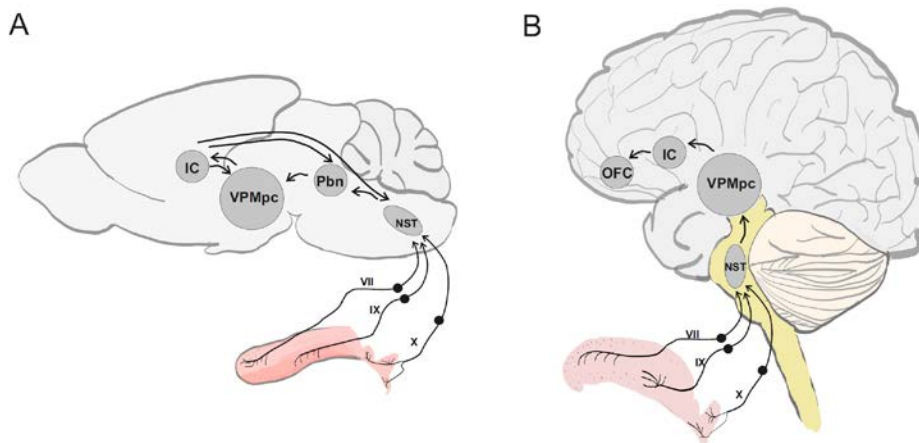
178 role from narrowly tuned TRCs as well as the contribution of broadly-tuned TRCs to coding of taste
179 signals remain, however, still unclear.

180 Taste quality coding begins with the sensitivities of individual receptor cells within taste buds. The
181 synaptic connections between these cells and gustatory nerve fibers is a major unknown at present.
182 Understanding convergence or divergence at these peripheral synapses will be key to understanding
183 the initial coding of taste signals in the periphery.

184

185 **Quality coding in the first neurons of the taste pathway**

186 How do primary sensory afferent neurons transmit taste information to the central nervous system (CNS;
187 see Figure 1) and how does activity in primary afferents represent taste quality (sweet, salty, sour, etc.)?



188

189 **Figure 1.** Schematics of the rodent (A) and human (B) gustatory pathways with a focus on peripheral and thalamo-
190 cortical relays. In both species, information is conveyed via cranial nerves VII, IX, and X from the tongue to the
191 brainstem. NST: nucleus of the solitary tract, Pbn: parabrachial nucleus, VPMpc: parvocellular portion of the
192 ventroposteromedial nucleus of the thalamus, IC: insular cortex, OFC: orbitofrontal cortex.

193

194 Electrophysiological recordings and Ca^{2+} imaging studies from primary sensory afferent neurons (single
195 fibers or ganglion neuron somata) have been carried by several groups. Some form of combinatorial
196 coding in taste was originally suggested by Pfaffmann (1941) based on early electrophysiological
197 recordings from afferent fibers that innervated taste buds in the cat. Single units were found that
198 responded to lingual stimulation with more than one taste compound (for example quinine or HCl or

199 both). That many fibers were not limited to excitation by a single taste quality, was inconsistent with a
200 labeled line coding scheme. This led Pfaffmann (ibid) to conclude "[...] sensory quality does not depend
201 simply on the "all or nothing" activation of some particular fiber group alone, but on the pattern of other
202 fibers active." Other investigators elaborated and extended this model to encompass the widespread
203 co-activation of a large number of sensory afferent fibers, with different combinations of the same fibers
204 constituting the code for different taste qualities. This was termed "cross-fiber coding" and was held as
205 the polar opposite of labeled line coding (Erickson 2008). According to cross-fiber coding, activity in any
206 single fiber on its own does not convey information about sweet, sour, salty, etc. Only the combined
207 activity of many fibers generates the code. Some resolution of these two opposite concepts—labeled
208 line versus combinatorial coding—was obtained by Frank and Pfaffmann (1969). They recorded from
209 single sensory afferent fibers from the tongues of hamsters and observed that although many fibers did
210 indeed respond to multiple taste stimuli, the most effective stimulus of a fiber was predictive of the
211 relative effectiveness of the other stimuli. These observations suggested that there were fiber "types"
212 organized according to the stimulus that evoked the "best" response. They termed these "sweet-best",
213 "salt-best" etc. fibers. Although this has been interpreted as a form of labeled line coding, the fact is that
214 activity in a single fiber could not unambiguously distinguish between (strong) excitation by the "best"
215 stimulus versus (weak) excitation by other, less effective stimuli.

216 The observation of "best stimulus" for individual taste afferent fibers has been widely replicated in
217 different laboratories, and in mammalian species ranging from mice to monkeys (Danilova *et al.* 1999;
218 Hellekant and Ninomiya 1994; Sato *et al.* 1975; Tonosaki and Beidler 1989). A further refinement of the
219 distinctions between taste afferents was the recognition that some neurons respond principally or
220 exclusively to one stimulus type, usually sugars – the so-called "specialist" neurons; other neurons
221 responded to a variety of electrolytes that might produce sour, bitter or salty tastes (reviewed by Frank
222 *et al.* 2008). Specialist and generalist neurons have been detected electrophysiologically as single-fiber
223 recordings on afferent nerves and by extracellular recordings in geniculate ganglia. A method applied
224 more recently is functional imaging of sensory afferent neuron activity using genetically encoded Ca²⁺
225 indicators such as GCaMP. Barretto *et al.* (2015) and Wu *et al.* (2015) carried out functional imaging on
226 geniculate ganglion neurons in the mouse and cataloged responses to a battery of different taste stimuli
227 presented at different concentrations. Those studies verified that about half the ganglion neurons were
228 "specialists" that responded best (and some solely) to a single taste compound, such as sucrose.
229 Specialist neurons could be detected for each of the five "basic" taste qualities (sweet, sour, salty, bitter,
230 umami). The geniculate ganglion also had "generalist" sensory neurons that responded much more
231 broadly to taste stimuli, mirroring the electrophysiological recordings from the primary afferent axons
232 (above).

233 The relative proportion of specialist and generalist neurons varied strongly depending on the
234 concentrations of stimuli tested (Wu *et al.* 2015). Importantly, neurons that displayed a specialist profile
235 with a low concentration stimulus were transformed to generalists when the same stimuli were tested at
236 higher concentrations. At concentrations that produced maximal responses, half the neurons exhibited
237 responses to multiple distinct stimuli. Unless half the information from the periphery is discarded, which
238 seems unlikely, a resolution to the question of taste coding is that a cross-fiber code involving a
239 combination of primary afferent axons that vary in their “tuning”, from specialists to generalists, encode
240 taste.

241 In addition to the encoding the basic taste qualities, there is a question of how stimuli which produce a
242 similar quality may be discriminated from one another. For instance, in primates, individual afferent fibers
243 that responded to one sweet stimulus typically also responded to several other sweets, and minimally
244 to bitter or sour tastants (Hellekant and Ninomiya 1994; Wang *et al.* 2009). This type of narrow tuning is
245 much less prevalent for taste qualities other than sweet: individual neurons respond quite variably to
246 different salts (Frank *et al.* 2008). However, this feature remains incompletely explored in the periphery
247 as most studies have utilized only limited panels of taste stimuli.

248 Whether sensory afferent fibers and their parent ganglion neurons employ patterns of action potentials
249 to encode stimulus identity has been explored to only a limited extent. Different taste stimuli appear to
250 cause primary afferent fibers to fire action potentials with somewhat different patterns, though these
251 differences are not marked (Lawhern *et al.* 2011; Nagai and Ueda 1981; Ogawa *et al.* 1974). Thus, spike
252 discharge pattern may augment and refine the combinatorial coding described above (Nagai and Ueda
253 1981). Taste coding in the periphery most likely involves activating a combination of afferent fibers
254 having varying tuning capabilities (from specialists to generalists) and subtly different firing patterns. All
255 these factors together play a role in the transmission of information needed to discriminate sweet, sour,
256 salty, bitter, and umami.

257 Parenthetically, a key point that should be noted is that to date, recordings from the primary afferent
258 neurons have only been obtained in anesthetized animals. It is possible that some of the distinctions
259 noted below in the response properties of higher level neurons may be attributable to anesthesia.

260

261 **Hindbrain neurons: evidence for temporal coding**

262 Gustatory afferents from the periphery project directly to the NTS in the brainstem where there is
263 substantial convergence (Whitehead and Frank, 1983; Whitehead, 1986). Cells in the brainstem, NTS
264 and PbN (the main target of projections from the NTS), are generally more broadly tuned than peripheral
265 fibers in both anesthetized (see Spector and Travers 2005, for a review) and awake (see Roussin *et al.*

266 2012, but see Nakamura and Norgren 1991) rodents, though there are still groups of neurons in each
 267 structure that are narrowly tuned to a single taste quality. Like fibers/cells in the periphery, neurons in
 268 the brainstem can become more broadly tuned with changes in stimulus concentration. Moreover,
 269 response profiles, defined as the subset of taste qualities that evokes a response, of NTS and PbN cells
 270 can change over time (Sammons et al., 2016). This may be due to the changing inputs to these cells as
 271 taste receptor cells die and are replaced. Despite such turnover, the network obviously needs to remain
 272 stable in its output. It is possible that extensive convergence from neurons with different profiles of
 273 sensitivities may support this stability; that is, the loss or addition of a few inputs with different taste
 274 sensitivities would have minimal impact on the target cells if there were enough variety in the array of
 275 inputs. Further, simultaneous recordings from taste-responsive NTS and PbN cells have shown that
 276 NTS with a particular best stimulus are more effective in driving PbN cells with a similar best stimulus,
 277 though the same PbN cells receive input from NTS cells with all types of best stimulus preferences (Di
 278 Lorenzo and Monroe, 1997; Di Lorenzo et al. 2009). As a changing array of inputs to NTS cells shift
 279 their response profiles from one best stimulus to another, simultaneous activation of enough inputs
 280 responding to a given best stimulus may also cause PbN cells upstream to shift their best stimulus in
 281 kind, as well as modifying the effectiveness of inputs that were activated. Thus, response profiles may
 282 change but the overall proportions of the constituents of the network encoding taste stimuli may remain
 283 consistent.

284 With a variety of response profiles in the taste-responsive portion of the NTS and PbN, there remains
 285 the problem of how confusion among similar-tasting, but not identical, tastants is resolved. As
 286 discussed, the across-fiber/neuron patterns may offer one solution, but another might be response
 287 dynamics, that is, temporal coding. Variation in the temporal pattern of taste-evoked firing offers a way
 288 to disambiguate two tastants that evoke similar response magnitudes within the same cell (Di Lorenzo
 289 et al. 2009).

290 Both specialist and generalist neurons have been described in brainstem taste areas in
 291 electrophysiological studies with anesthetized animals. Perceptually similar stimuli evoke similar
 292 patterns of neuronal population activity, lending support to the combinatorial coding model discussed
 293 above (Geran and Travers 2009; Simon *et al.* 2006; Smith *et al.* 2000). However, unlike taste bud cells
 294 and sensory afferent neurons, gustatory neurons of the brainstem do exhibit evidence of temporal
 295 coding. "Metric space analysis" (MSA; Victor and Purpura 1996; 1997) has been used to quantify this.
 296 MSA begins by determining a "distance" between spike trains in terms of the "cost" of making them
 297 identical, via adding, deleting, or moving spikes. Adding or removing a spike costs one arbitrary unit.
 298 The cost of moving a spike in time by an amount t is given by qt , where q is a parameter that controls
 299 the sensitivity of the distance to spike timing. Based on these distances, calculated from repeated neural

300 responses to presentations of several tastants, one can determine two information-theoretic quantities:
301 H^{count} and $H^{\text{spike}}[q]$. H^{count} is the amount of information about taste quality conveyed by spike count alone,
302 and $H^{\text{spike}}[q]$ is the amount of information about taste quality when spike timing is taken into account.

303 In early work using anesthetized rats, spike timing was shown to convey a significant amount of
304 information about taste stimuli in both the Nucleus of the Solitary Tract (NTS; Di Lorenzo and Victor
305 2003) and the Parabrachial Nucleus of the pons (PbN; (Rosen *et al.* 2011), respectively the first and
306 second synapses in the central gustatory pathway in rodents. Specifically, in about half of the taste-
307 responsive cells in NTS (Di Lorenzo and Victor 2003) and PbN (Rosen *et al.* 2011), spike timing
308 contributes to taste quality discrimination – and in both NTS and PbN this contribution was largest in
309 neurons that would appear to be broadly tuned if only spike count were considered. In addition, in the
310 NTS, spike timing contributes significant amounts of information to distinguishing among responses to
311 the components of binary mixtures (Di Lorenzo *et al.* 2009b), between tastants of different
312 concentrations (Chen *et al.* 2011a) and tastants of the same taste quality but different chemical
313 compositions (Roussin *et al.* 2008).

314 While evidence for temporal coding of taste stimuli in brainstem structures has been obtained in the
315 anesthetized animal, further studies asked whether there was similar evidence of temporal coding of
316 taste in the alert animal (Roussin *et al.* 2012; Weiss and Di Lorenzo 2012). To that end, rats were
317 implanted with 8-channel microwire electrode bundles aimed at either the NTS or PbN. Following
318 recovery from surgery, mildly water-deprived rats were placed in an experimental chamber with a
319 drinking spout that allowed control of various fluids on a lick-by-lick basis. Taste responses in the NTS
320 and PbN of awake freely licking rats differed in several ways from those recorded under anesthesia. For
321 example, in addition to the typical phasic-tonic time course of response seen under anesthesia, brief
322 lick-by-lick responses were also apparent in many NTS and PbN cells recorded in awake rats. Of these,
323 some cells had responses that progressively increased with successive licks. There were also many
324 cells with very long latency (>2 sec) taste responses that began long after the licks were completed
325 (Roussin *et al.* 2012), which might be the result of stimulation of post-oral receptors during swallowing.

326 Recordings from the NTS (Roussin *et al.* 2012) and PbN (Weiss *et al.* 2014) of awake freely licking rats
327 revealed a rich variety of cell types in addition to those that respond solely to taste. For example, many
328 cells fire in phase with licking, with peak firing rates just at the time of the lick, or between licks. In
329 addition, there are cells that significantly decrease their firing rate during a lick bout. The relative
330 silencing of such cells when the rat engages in consummatory behavior suggests that they may set the
331 initial conditions for the network to acquire sensory information. Moreover, these data underscore the
332 idea that sensory and motor components of gustation are inextricably linked.

333 In a separate series of experiments, the effects of pairing olfactory stimuli with tastants were tested
334 (Escanilla *et al.* 2015). Widespread modulation of taste responses was observed, including changes in
335 response magnitude and latency following taste-odor pairing. MSA of taste- and odor-evoked responses
336 showed that NTS cells were more competent at discriminating tastants when they were presented with
337 odors than when presented alone. This applied for all taste qualities, and whether or not spike timing
338 was taken into account, leading to the hypothesis that brainstem neurons may be most keenly tuned to
339 respond to naturalistic stimuli, that is food, rather than pure chemical exemplars of taste qualities
340 (Escanilla *et al.* 2015). This was tested by presenting complex, natural stimuli such as grape juice
341 (sweet), clam juice (salty), lemon juice (sour) and coffee (bitter). Evoked spike trains in the PbN of awake
342 freely licking rats displayed conveyed significantly more information to naturalistic stimuli than those
343 associated with single compounds (Weiss *et al.* 2014).

344 In conclusion, data from electrophysiological recordings from awake, freely licking rats, underscores the
345 role of the gustatory brainstem as an important node in the neural circuit that controls food identification
346 and ingestion. In addition, dynamics – both intrinsic to the spike trains and related to the lick cycle – are
347 prominent and functionally significant aspects of neural responses.

348 From the gustatory brainstem, afferents target the most medial portion of the ventral posteromedial
349 thalamus. Taste-responsive thalamic neurons in this nucleus form an important source of input to
350 gustatory cortex. Although this small region has been understudied relative to other taste areas, there
351 is recent evidence that the gustatory thalamus may play important roles in taste quality and palatability
352 coding, as well as stimulus expectation (Liu and Fontanini 2015).

353

354 **Patterns of activity in the rodent gustatory cortex**

355 Within gustatory cortex (GC), physiological studies demonstrate that taste-responsive cells are often
356 multimodal, responding to other sensory stimuli in addition to taste (for review, see Maffei *et al.* 2012).
357 When recordings are made in either anesthetized or awake animals probed with only sapid stimuli,
358 both narrowly and broad taste-responsive neurons are found, similar to those found in both peripheral
359 and other central taste areas (e.g. Katz *et al.* 2001; Ogawa *et al.* 1992a; Ogawa *et al.* 1992b; Sadacca
360 *et al.* 2016; Spector and Travers 2005; Stapleton *et al.* 2006a; Yamamoto *et al.* 1989; Yamamoto *et al.*
361 1984; Yamamoto *et al.* 1985). The roles of these cell types are still ambiguous in terms of function,
362 although there is evidence that some cortical taste neurons may respond broadly to sets of stimuli that
363 can be classified as sharing a hedonic value (Fontanini and Katz 2006; Yamamoto *et al.* 1989).

364 An important and related, yet not well-understood aspect of taste coding, involves the spatial
365 organization of taste neurons – are cells responsive to particular taste stimuli clustered together? Other

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367 sensory systems differ in this mode of organization; from the well-known somatotopy of barrel cortex, to
368 the apparent random overlap of odorant responses in piriform cortex (Petersen 2007; Stettler and Axel
369 2009). Chen and colleagues (2011b) used 2-photon imaging to describe a sharply segregated quality
370 representation in mouse GC. Here, quality-specific clusters of singly responsive neurons were separated
371 in space along the cortical surface, by areas with only sparse taste-evoked activity. In contrast, the vast
372 majority of work on taste cortex is entirely consistent in suggesting that there is little to no stimulus
373 topography in how taste qualities are represented in the gustatory cortex. Across the anterior – posterior
374 expanse of GC, mapping studies using different techniques have yielded very different conclusions. For
375 instance, studies using either in vivo recordings, or intrinsic imaging, show a large degree of overlap
376 among the basic taste stimuli, with bias towards overrepresentation of individual qualities at the anterior
377 and posterior extremes (Accolla *et al.* 2007; Bahar *et al.* 2004; Carleton *et al.* 2010; Yamamoto *et al.*
378 1985). A genetically encoded trans-synaptic tracer similarly suggested that neurons receiving input for
379 different taste qualities are intermingled in the gustatory cortex (Sugita and Shiba 2005).

380 More recently, 2-photon imaging was used to investigate taste responses to stimuli representing four
381 primary qualities (acid, bitter, salty and sweet) in an area of mouse gustatory cortex defined by taste
382 thalamic input (Fletcher *et al.* 2017). This “central” area, located just posterior to the landmark middle
383 cerebral artery, possessed thalamic terminal labeling concentrated in the dysgranular subdivision. Using
384 a virally expressed calcium indicator (GCaMP6s), taste imaging responses were collected in
385 anesthetized mice in this delineated area. Not surprisingly, cortical taste cells were found to respond
386 either best to individual stimuli, or combinations of stimuli. Spatial mapping demonstrated that taste
387 quality responses overlapped in this region, with no evidence of segregation of cells responding to a
388 single quality. Principle components analysis of this aggregate data suggested that the primary taste
389 qualities were distinctly represented in the population response, providing a basis for discrimination
390 despite spatial overlap. Moreover, the stimuli were ordered along the first component in a way that
391 suggested hedonic character may also be represented in the response.

392 The finding of an area of quality overlap in the center of mouse GC fits in nicely with the previously
393 mentioned mapping studies in the rat (Accolla *et al.* 2007; Yamamoto *et al.* 1985), and other recent 2-
394 photon approaches in mice (Lavi *et al.* 2018; Livneh *et al.* 2017). Still, these papers and the Chen *et al.*
395 (2011b) study leave open the possibility that bitter taste responses and sweet taste responses may be
396 overrepresented posteriorly and anteriorly, respectively, in GC. If so, any topographic representation of
397 taste quality likely stems from the source of peripheral input. The glossopharyngeal (IX) nerve, which
398 innervates posterior taste buds, is known to be more responsive to bitter-tasting stimuli than branches
399 of the facial nerve (VII), which innervate taste buds on the anterior tongue and palate (Frank 1991; Frank
400 *et al.* 1983). In rat taste cortex, information from the chorda tympani branch of VII projects to the anterior

GC, while information from IX targets the posterior GC (Hanamori *et al.* 1998; Yamamoto *et al.* 1980). A similar “gradient” of taste quality representation that follows peripheral input has also been described in the parabrachial nucleus in the rodent brainstem (Geran and Travers 2006; Halsell and Travers 1997). In this discussion, however, it is important to consider the multimodal nature of GC, as well as surrounding cortical areas. For example, there is also a prominent viscerosensory representation found in posterior insular cortex, adjacent to GC (Cechetto and Saper 1987). Perhaps correspondingly, the hotspot for conditioned taste aversion learning is also found in posterior insular or GC (Schier *et al.* 2016; Schier *et al.* 2014). Furthermore, Hanamori and colleagues (1998) found that over 75% of taste-responsive neurons in posterior GC in rat were also responsive to a nociceptive stimulus (tail pinch).

In summary, reports (Chen *et al.* 2011b; Peng *et al.* 2015) from a single laboratory notwithstanding, the evidence is now quite strong that gustatory signals for taste quality are distributed and intermingled in the gustatory cortex.

Patterns of gustatory activity in the human cortex

While taste processing in the periphery and also the central nervous system has gained considerable attention in animal models, these processes are still to be investigated in humans. Of particular interest are questions on how, when, and where taste information, in general, and specific taste attributes such as taste quality, intensity, and hedonics, in particular, are processed in the human brain. Human neuroimaging studies have shown that taste consistently activates a range of cortical areas including the anterior insula and frontal operculum (FOP), mid-dorsal insula and overlying Rolandic operculum, posterior insula and POP, as well as the postcentral gyrus (cf. Veldhuizen *et al.* 2011; Yeung *et al.* 2018). Evidence suggests that the mid-dorsal insula and the adjacent FOP form GC (Bender *et al.* 2009; Iannilli *et al.* 2012; O'Doherty *et al.* 2001; Small 2010; Small *et al.* 1999), while the posterior insula and POP have been implicated in oral somatosensation and attention to the mouth rather than gustation (Veldhuizen *et al.* 2007). These findings are in line with macaque anatomy, where the anterior and mid insula and the FOP, but not the POP, receive taste afferents from the thalamus (Pritchard *et al.* 1986) but may not directly translate to human physiology. Observations that taste sensations can be elicited by electrical stimulation of the mid-dorsal insula (Mazzola *et al.* 2017) further corroborate its role as GC. Consistent with the anatomical evidence, scalp-level electrophysiological studies found pronounced activation of the bilateral anterior in mid insula and adjacent frontal operculum in response to electric (Ohla *et al.* 2010) and sapid taste (Crouzet *et al.* 2015; Tzieropoulos *et al.* 2013) within 150 ms of taste delivery.

Functionally, insular activation has been linked with sensory taste features, such as taste intensity (Grabenhorst and Rolls 2008; Guest *et al.* 2007; Ohla *et al.* 2010; Spetter *et al.* 2010; Tzieropoulos *et*

434 *al.* 2013) and taste quality (Crouzet *et al.* 2015; Schoenfeld *et al.* 2004); taste pleasantness and
435 valuation, on the other hand, have been mostly associated with activity in the OFC, the anatomically
436 later, secondary taste area (Grabenhorst and Rolls 2008; Guest *et al.* 2007). However, it has also been
437 proposed that the GC jointly encodes both the chemical identity and palatability of a tastant (de Araujo
438 *et al.* 2006) thereby suggesting a role of the insula in the evaluation of taste or its precursors beyond
439 mere sensory processing. This notion is corroborated by observations that expectations about the value
440 of a taste, induced by visual cues, modulate taste-related processing in the rodent (Grossman *et al.*
441 2008) and in the human (Nitschke *et al.* 2006; Ohla *et al.* 2012) insula.

442 In contrast to animal models, the mechanisms underlying taste quality coding have received little
443 attention in humans mostly due to the limited spatial resolution of noninvasive brain imaging techniques
444 such as functional magnetic resonance imaging (fMRI) yielding a spatial resolution of a few millimeters
445 at best. Accordingly, only a few fMRI studies have addressed the question of a gustotopic organization
446 of the human GC and their results failed to provide evidence for a clear spatial segregation of taste
447 qualities but rather suggest a partial overlap of insular representations for different tastes (Dalenberg *et al.*
448 2015; Prinster *et al.* 2017; Schoenfeld *et al.* 2004). However, cortical activation patterns change
449 rapidly, within milliseconds, rendering temporal information a candidate variable for taste quality coding.
450 In fact, neuronal response patterns obtained from electrophysiological recordings at the scalp allow
451 deciphering which taste participants tasted on a given trial. The onset of this discriminability coincided
452 with the earliest taste-evoked responses that were localized in GC signifying that quality is among the
453 first attributes of a taste represented in the central gustatory system (Crouzet *et al.* 2015) in strong
454 accord with electrophysiological studies in awake rodents (Graham *et al.* 2014; Pavao *et al.* 2014;
455 Stapleton *et al.* 2006b). The results also align with and add to observations that neuronal response
456 patterns along the rodent gustatory neuroaxis, including the nucleus of the solitary tract (Di Lorenzo *et al.*
457 2009a), parabrachial nucleus (Geran and Travers 2013), and insula (Jezzini *et al.* 2013), code taste
458 quality.

459 More recent evidence linked the predictive value of gustatory neural response patterns and taste-related
460 decision-making. For this, behavioral reports from different tasks were combined with multivariate
461 analyses of large-scale electrophysiological recordings in a series of studies. Specifically, Crouzet and
462 co-workers (2015) showed that the more alike the neural response patterns of any two tastes were, as
463 indicated by poorer discriminative performance of a classifier, the more these tastes were confused by
464 the participants. The results were surprising for the taste domain because they provide evidence for a
465 mapping between neural and phenomenological rather than between neural and chemical spaces.
466 Whether the information encoded in gustatory neural response patterns drives actual behavior was
467 addressed in two further studies. In the study by Wallroth and Ohla (2018), participants were to detect

the presence of a taste as quick as possible. They found that the onset of taste decoding (discriminable brain response patterns) indeed predicted *when* participants detected a given taste by button press and linked neuronal response patterns to the speed of simple gustatory perceptual decisions – a vital performance index of nutrient sensing. Interestingly, the onset of taste decoding was earlier in this study, where participants responded speedily, compared to the previous study, where participants performed a delayed response task suggesting that the timing of gustatory coding is in a way flexible and dependent on behavioral goals.

While the mere detection of a taste in the oral cavity may prepare a non-specific response, the regulation of nutrient uptake and expulsion of potential toxins calls for quick and reliable taste detection and identification. Whether taste detection and discrimination are sequential or parallel processes, that is whether you know what it is as soon as you taste it, was addressed in another study (Wallroth and Ohla *in press*). To uncover the sequence of processing steps involved in taste perceptual decisions, participants performed taste-detection and -discrimination tasks. Irrespective of taste quality and task, neural decoding onset and behavioral response times were strongly linked, demonstrating that differences between taste judgments are reflected early during chemosensory encoding. Moreover, neural and behavioral detection times were faster for the iso-hedonic salty and sour tastes than their discrimination time. No such latency difference was observed for sweet and bitter, which differ hedonically. These results indicate that the human gustatory system detects a taste faster than it discriminates between tastes, yet hedonic computations may run in parallel (Perez *et al.* 2013) and facilitate taste identification.

Together these studies clearly show that the information encoded in taste-related neural response patterns is also the foundation for gustatory decision-making and that the timing aligns with task-specific goals.

Cortical population coding of taste decisions and behavior

Taste quality is tightly linked to taste palatability or pleasantness. While sweet taste is typically liked, bitter taste is commonly aversive to most mammals. Accordingly, the gustatory neuroaxis needs to represent both features as they, together, drive food-related decisions and allow adaptive behavior. In awake rats, taste administration is represented by complex temporal coding in single neurons: a brief period of non-specific firing is followed by approximately 500 msec of identity-related firing, which is in turn replaced by firing that is reliably related to taste palatability (Katz *et al.* 2000; Sadacca *et al.* 2012). A series of studies have demonstrated that the palatability “epoch” can be independently manipulated, validating the characterization: changes in perceived palatability, such as that observed at the transition

501 from an attentive to “withdrawn” state (Fontanini and Katz 2005; 2006) and across conditioned taste
502 aversion learning (Grossman *et al.* 2008; Moran and Katz 2014), change palatability epoch coding while
503 having no impact on the earlier ~800 ms of taste-induced activity.

504 CNS neural responses provide information about the identity of tastes on the tongue. Countless studies
505 have demonstrated that sapid stimuli, flowing across the tongue of anesthetized animals, induce
506 responses in neurons across the gustatory neuroaxis (for just a few examples, see (Azuma *et al.* 1984;
507 Di Lorenzo 1988; Di Lorenzo and Victor 2003; Erickson *et al.* 1994; Li *et al.* 2013; Nishijo and Norgren
508 1990; Yamamoto 1984; Yamamoto *et al.* 1989). Perhaps the most discussed facet of these studies is
509 the fact that taste responses vary vastly in breadth; a great deal of energy has been devoted to debating
510 theories of gustatory coding that turn on these breadths of responsivity (e.g., Di Lorenzo 2000; Lemon
511 and Katz 2007; Scott 2004; Smith and St John 1999; Spector and Travers 2005). Neural circuitry in
512 general, and taste circuits in particular, are rife with cross-talk and feedback at both micro- (within region)
513 and macro-circuit (between region) level (Jones *et al.* 2006). Empirical and theoretical work makes it
514 clear that neural responses in such interactive networks should contain functionally interpretable
515 dynamics that are most meaningful when examined at the ensemble rather than at the single cell level
516 (e.g., see Abarbanel and Rabinovich 2001).

517 An independent set of studies have made use of analytic techniques specialized to interpret the real-
518 time firing of multiple simultaneously-recorded neurons (hidden Markov modeling, or HMM). This work
519 reveals that firing rate modulations within taste responses, which appear gradual in across-trial averages
520 of single-neuron firing, are in fact not gradual at all. Rather, they reflect sudden coherent shifts between
521 ensemble states: at particular time points within individual trials, the firing rates of (on average) ~50% of
522 the recorded neurons will change simultaneously; across-trial averages “smear” these changes, making
523 them look more gradual, because they happen at different latencies in different trials (Jones *et al.* 2007;
524 Miller and Katz 2010). Together, these two sets of results suggest the testable hypothesis that GC neural
525 ensembles, far from simply coding what the taste IS, may process that information to directly drive action.
526 If in fact palatability-related firing appears suddenly in single trials (a possibility implied by but not directly
527 demonstrated in the above-described work), it is possible to hypothesize that this appearance predicts
528 the onset of consumption behavior. Our testing (Sadacca *et al.* 2016) proves this to be the case, in that
529 analyses keyed to the onset of the ensemble state dominant during the palatability epoch (rather than
530 to stimulus onset time, as is more typical) reveal that palatability coding does emerge suddenly—more
531 suddenly than a range of ramping models (including the model used to explain primate perceptual
532 decision-making (see Gold and Shadlen 2001; Shadlen *et al.* 1996) can explain, and as fast as models
533 assuming instantaneous state transitions (Sadacca *et al.* 2016).

534 Armed with the knowledge of precisely *when* decision-related information appears in GC on individual
535 trials, the authors were then able to compare this information to within-trial latencies of palatability-
536 related behavioral responses, measured through electromyography. This analysis specifically reveals
537 that the sudden emergence of the “palatability-related state” in GC neural ensembles predicts both
538 *whether* the rat will gape in response to taste stimulation and precisely *when* that gape will occur, in
539 single trials, with correlation values of ~ 0.75 (Sadacca *et al.* 2016).

540 The above results, while robust, are phenomenological. Li and co-workers (2016) performed two types
541 of perturbation experiments to test whether GC ensemble transitions are causally linked to consumption
542 behavior. In one experiment, arrival of an aversive taste was cued: as the rats learned the meaning of
543 the cue across a full session, the latency with which they gaped in response to the taste decreased by
544 ~ 150 ms; recordings showed that the cue had an almost identical impact on neural coding of that
545 aversive taste. In the second experiment; optogenetic silencing of GC neurons was shown to change
546 the likelihood of gaping. Together, these experiments confirm the general hypothesis that GC is a part
547 of a distributed system responsible for transforming an incoming identity code into a taste decision.

548 These results, while perhaps surprising within the field of taste research, are consistent with a great deal
549 of work on sensorimotor systems—and, more specifically, on work describing the top-down modulation
550 of multi-rhythmic central pattern generators (Marder 2012).

551

552 **Conclusion**

553 When examined at each level of the nervous system – periphery, brainstem, and cortex – it is evident
554 that individual taste-responsive receptor cells or neurons may respond either selectively or broadly to
555 stimuli of different taste qualities. Recent approaches to rodent and human central taste also emphasize
556 the importance of temporal response patterns, which likely underlie the progression of taste behavior,
557 from detectability to discrimination. This response complexity supports the notion of combinatorial
558 coding along the gustatory neuroaxis. The flexibility inherent in this type of coding for the sense of taste
559 may be necessary for animals to exhibit adaptive behavior in food selection and consummatory
560 behavior.

561

562 **Acknowledgement**

563 The authors acknowledge the following sources of support: BMBF 01EA1408A-G (K.O.),
564 R01DC014420, and R21DC012746 (S.D.R. and N.C.), R01DC007630 (S.D.R.), R01DC016833 (J.B.
565 and M.F.), R01DC006914 (P.M.D.), R01DC00945, R01DC007708, and R01DC006666 (D.B.K.),

566 R01DC006304 (N.C.), JSPS KAKENHI JP26462815, 18K09507 (R.Y.). The authors thank Iryna Ruda
567 for help with the artwork in Figure 1.

568

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